

METHOD FOR THE REMOVAL OF ALOIN, EMODIN AND/OR
ISO-EMODIN FROM ALOE VERA BY TREATMENT WITH AN OXIDASE

The present invention relates to a method of removing
5 aloin, emodin and/or iso-emodin from Aloe Vera gel and
to a method of preparing Aloe Vera gel with a content
of less than 5 ppm of aloin, emodin and/or iso-emodin,
where manual filleting of the Aloe Vera leaves and
losses in relation to conventional preparation methods
10 are avoided.

In the Aloe Vera plant family, there are the two
species *Aloe barbadensis* Miller and *Aloe aborens*,
both of which are currently exploited commercially.
15 These perennial plants from the family *Liliaceae* have
their origin in Eastern Africa; they are currently
grown essentially along the border between the United
States and Mexico. Cultivation in Europe is
insignificant.

20 Two products can be obtained from Aloe Vera leaves. The
region directly underneath the cortex gives a yellowish
juice (a type of latex). This juice contains aloin, a
glycosylated anthrachinon derivative, as its main
25 constituent (see fig. 1). Aloin is a strong laxative
which has been used as such for more than 2000 years;
nowadays, it is unimportant. The inner tissue of the
plant's leaves, which has a high water content, gives
the Aloe Vera gel, which is employed in a number of
30 ways in cosmetics and other natural products. Its
annual turnover is estimated to be 7 billion €. This
Aloe Vera gel must no longer contain any aloin, or the
unglycosylated anthraquinone derivatives emodin or iso-
emodin. A content of less than 5 ppm is desired, but
35 not achieved with all products. In the prior art, Aloe
Vera gels with aloin contents of less than 20 ppm have
hitherto only been achieved by laborious manual
filleting of the Aloe Vera leaves. In the prior art,

Aloe Vera products with low aloin contents always involve losses some of which are considerable.

5 The processing into Aloe Vera gel usually involves the following procedure. After washing, the leaves are filleted (Fig. 2) in order to remove the aloin before further processing. As a result of the concave shape of the leaves, their substantial size differences and the structure of the leaf, this filleting step is
10 predominantly carried out manually. This procedure results in losses of 20 - 60% of the harvested material.

15 In the past there has been no lack of attempts to peel the Aloe Vera leaves mechanically; however, the aloin concentration which remains in the product is unacceptably high.

20 It was therefore the object of the present invention to provide a method with which the aloin concentration in products such as the Aloe Vera gel can be lowered substantially and manual filleting can be avoided, without losing an unduly high proportion of harvested material in the process.

25 This object was solved in accordance with the invention by using oxidases.

30 Nothing is known from the relevant specialist literature about enzymes or enzyme systems which have a specific aloin-degrading activity; however, in the gastrointestinal tract, aloin is deglucosidated by glucosidases, and the resulting iso-emodin has the actual laxative activity.

35 A qualitative detection of aloin, emodin and their isomers can be performed particularly successfully by thin-layer chromatography. A suitable quantitative

analytical method is in particular photometry, because the compounds in question absorb highly in the visible light range. However, none of the methods allow any conclusions regarding the nature of any degradation products which may have formed. These were analyzed by gas chromatography coupled with mass spectrometry (GC-MS). However, only emodin (which is unglucosidated) can be vaporized without decomposition and is therefore suitable for GC analysis.

Surprisingly, aloin has now been successfully degraded for the first time with the aid of oxidizing enzymes (oxidases), in particular from the classes of the peroxidases (E.C. 1.11.1.7) and laccases (E.C. 1.10.3.2.). As is known, peroxidases act on substrates such as, for example, guaiacol; laccases act on the lignin constituents; it must be considered as surprising that these enzymes accept aloin or emodin as substrates. The oxidant which was preferably used in the laboratory for this purpose was hydrogen peroxide, of which even small amounts can be metered precisely; in the case of larger batches, however, the oxidation can also be accomplished with the aid of atmospheric oxygen.

The degradation of aloin was first detected by thin-layer chromatography in direct comparison with an untreated sample. Moreover, a drastic drop in the typical absorption was observed in the photometer at 328 nm. It has been observed that the degradation of emodin is completely analogous. Moreover, after emodin has been subjected to oxidative degradation, a gas chromatograph shows four new signals at a markedly reduced retention time. The evaluation of the mass spectra demonstrated that the signals were salicylic acid derivatives.

The invention therefore relates to a process for removing aloin, emodin and/or iso-emodin from Aloe Vera gel where the gel is brought into contact with an oxidase under conditions which are suitable for the enzymatic activity.

Thus, there is furthermore provided a process of preparing Aloe Vera gel with a content of less than 5 ppm of aloin, emodin and/or iso-emodin without manual filleting, where the gel is brought into contact with an oxidase under conditions which are suitable for the enzymatic activity. If appropriate, the oxidase is removed from the gel after the reaction has taken place, for example in order to re-use the enzyme.

According to a particular embodiment of the invention, the oxidase is a peroxidase, preferably peroxidase E.C. 1.11.1.7 from *Glycine max.*, or a laccase, preferably oxidase E.C. 1.10.3.2 from *Rhus vernificera*.

The oxidase can be present in isolated or purified form or else in the form of an extract from a natural substance. In the method according to the invention, the oxidant used is hydrogen peroxide or (atmospheric) oxygen.

The enzymatic reaction is advantageously carried out in an aqueous suspension or solution of the Aloe Vera gel. In accordance with a particular embodiment of the invention, the suspension or solution is buffered to the pH value which is the activity maximum of the enzyme.

The Aloe Vera gel is preferably obtainable by an abovementioned process, or process described in the examples section, respectively. However, it is clear to a person skilled in the art that modifications of the process according to the invention are possible (such

as, for example, variations of the reaction conditions, solvents, enzymes and the like), which essentially lead to the same or an improved result, which is the degradation of aloin, emodin and/or iso-emodin in Aloe Vera gel.

The particular advantage of the present invention is that laborious manual cutting up of the Aloe Vera leaves is not required and that therefore losses of the raw material are avoided and a very pure product is obtained.

Aloe Vera gel contains a large number of organic valuable constituents. Approximately 160 such constituents are known; the most important is what is known as alloverose, a pentasaccharide. Alloverose is virtually completely eliminated for example by the known unselective adsorption onto activated charcoal, with the aid of which it is also possible to reduce the aloin content (cf., for example, US 5,356,811), and the dermatological value of such Aloe Vera gels is thus distinctively reduced. In contrast, the method according to the invention for the biocatalytic removal of aloin does not attack alloverose.

The invention is illustrated in greater detail by the examples which follow; they, however, do not limit the invention described.

Examples

Example 1

0.5 mg of peroxidase (E.C. 1.11.1.7; *Glycine max.*) and 8 µl of a 3% hydrogen peroxide solution are added to 3 ml of an ethanolic solution of 1000 ppm of aloin in a 1cm PMMA cuvette. Within 2 hours, the absorbance in the photometer at 328 nm drops from 1.589 to 0.538. After

12 hours, the thin-layer chromatogram no longer reveals an aloin signal.

Example 2

5 0.5 mg of peroxidase and 8 µl of a 3% hydrogen peroxide solution are added to 3 ml of a solution of 200 ppm of emodin in tetrahydrofuran in a 1cm quartz glass
cuvette. Within 2 hours, the absorbance in the
photometer at 328 nm drops from 2.646 to 0.709. After
10 12 hours, the thin-layer chromatogram no longer reveals an emodin signal. The gas chromatogram reveals a new signal at 13.6 min (emodin 19.4 min). The mass spectrum shows dimer signals of the degradation products at 148 + 133 u.

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Example 3

Example 1 is repeated with an aqueous suspension of 1000 ppm of aloin. Again, the absorbance in the photometer at 328 nm drops, and the thin-layer
20 chromatogram no longer reveals an aloin signal after 12 hours.

Example 4

2 mg of peroxidase are added to 4 ml of a 2.5% solution
25 of emodin in ethanol/water (20/80 v/v) and a gentle stream of compressed air is subsequently passed in over two hours, using a Pasteur pipette. During this process, the color of the solution changes from deep yellow to dark red. A fraction with the characteristics
30 described in example 2 can be separated by column chromatography.

Example 5

Example 1 is repeated with 0.5 mg of a laccase (E.C.
35 1.10.3.2.; *Rhus vernificera*) whose activity had previously been ensured with syringaldazin. A suspension of 1000 ppm of aloin in phosphate buffer (pH value 6.5) with addition of 5% by volume of ethanol is

used. Again, the absorbance in the photometer at 328 nm drops, and the thin-layer chromatogram no longer reveals an aloin signal after 12 hours.

5 Example 6

Example 1 is repeated with an Aloe Vera gel (1:100) with an aloin content of 250 ppm (noncommercial product). Again, the absorbance in the photometer at 328 nm drops, and the thin-layer chromatogram no longer
10 reveals an aloin signal after 12 hours.

Example 7

Example 6 is repeated with an Aloe Vera gel (1:100) with a known alloverose content. The above-described
15 treatment does not reduce the alloverose content. If, in contrast, the Aloe Vera gel is filtered through activated charcoal, alloverose is no longer detectable thereafter.